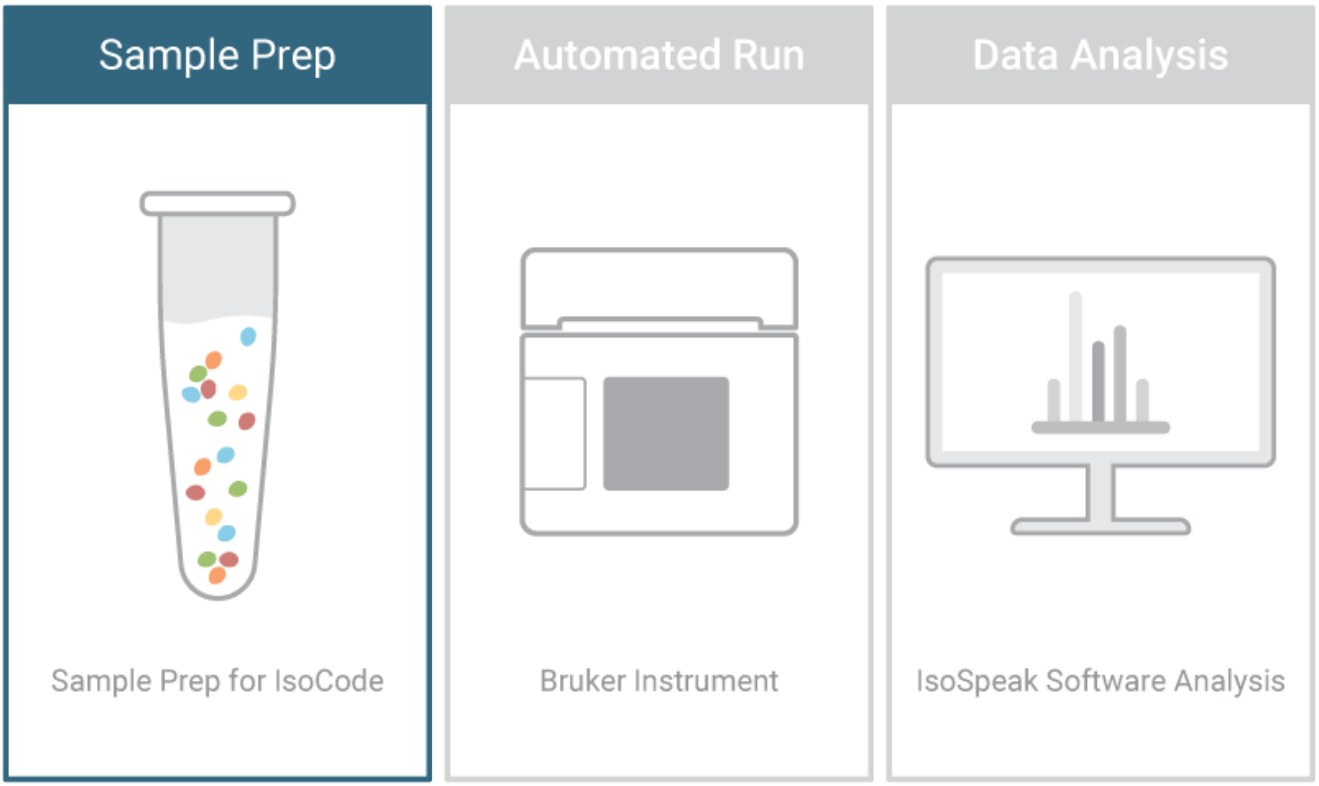
# IsoCode Single-Cell Adaptive Immune: Mouse T-cells Protocol

Ensure you achieve the maximum benefit from the Bruker systems and generate impactful data as quickly as possible






Key: ● TIP, ● CRITICAL, ● OPTIONAL

## Contents





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# A. Overview

## Overview of Protocol

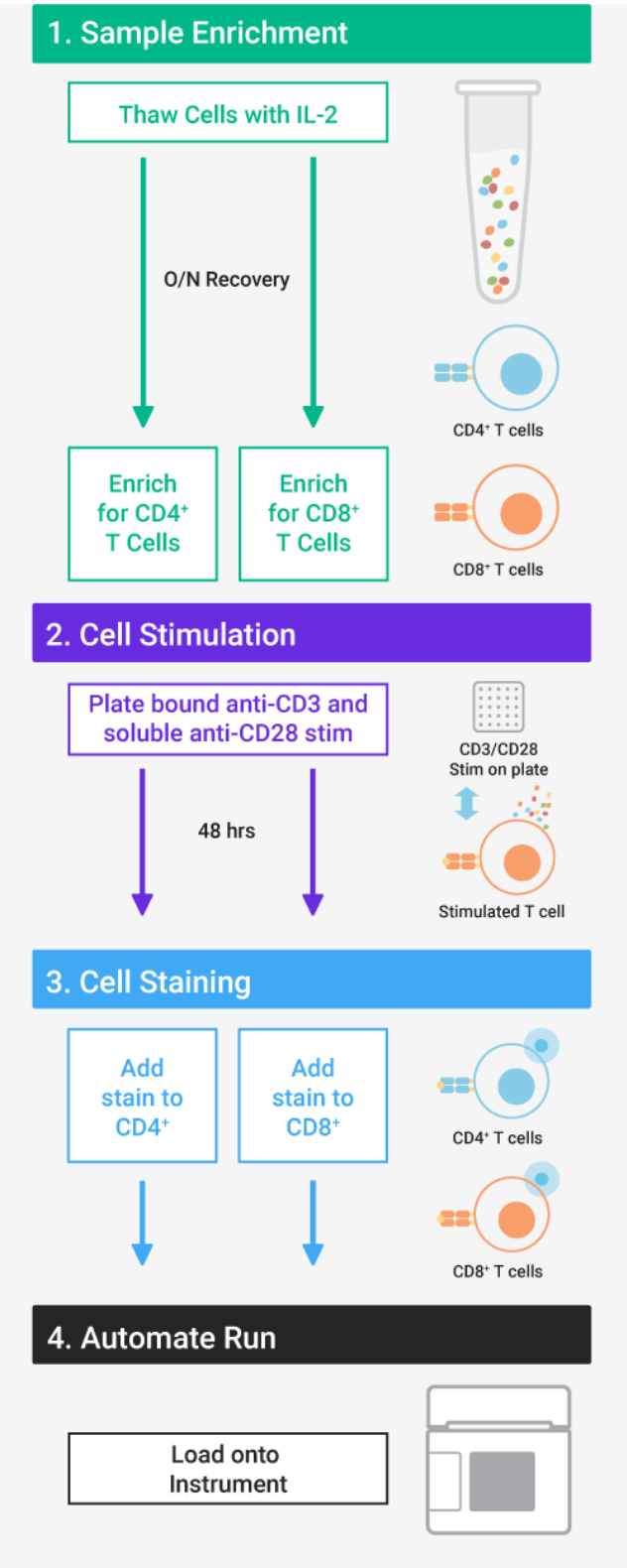
Day 1: Cryopreserved cells are thawed and cultured overnight in the presence of IL-2.

Day 2: Enrichment and Stimulation of CD4+ and/or CD8+ T cells for 48 hours.

Day 4: Staining and Loading of T cells onto IsoCode chip.

NOTE:
This protocol outlines the standard method for thawing and culturing of mouse T cells only and may not be valid for other species or cell types.

NOTE:
Using stains and protocols other than the included kit surface stains and protocols might result in failed runs. Stains and staining procedures not approved by Bruker will require validation prior to use. Please consider Bruker's IsoPACE program to assist in custom marker and protocol validation.




## Safety Warnings



Key: ● TIP, ● CRITICAL, ● OPTIONAL
PRO-7 REV 6.0

- Read MSDS documents of all materials prior to use.

- Laboratory workers should wear standard PPE, including disposable gloves, protective eyewear, and laboratory coats.

## Required Reagents, Consumables and Equipment

### Table 1: Required Reagents and Consumables Provided by Bruker

| Item | Catalog Number | Quantity | Comment |
|---|---|---|---|
| IsoCode Kit | Please see website (https://brukercellularanalysis.com/) for available kits or talk to Bruker's Customer Service team for details | One chip per sample/cell type/condition | Subcomponents stored at 4°C and -20°C |

### IsoCode Kit Components

IsoLight IsoCode Reagent Box (4°C)

- 15 mL Tube A
- 15 mL Tube B
- 1.5 mL Tubes A/B: Cocktail A in Micro-Tube (Green Cap) and Cocktail B in Micro-Tube (Red Cap)
- 50 mL Tubes containing Reagents 1, 2, 3, 4, 5, 6, 7, 8
- 1 Bag of Disposable Reagent Sippers
- Alexa Fluor 647 anti-mouse CD4 stain (AF647-CD4) [ordered separately]
- Alexa Fluor 647 anti-mouse CD8a stain (AF647-CD8) [ordered separately]

IsoSpark IsoCode Reagent Box (4°C)

- 15 mL Tube A
- 15 mL Tube B
- 1.5 mL Tubes A/B: Cocktail A in Micro-Tube (Green Cap) and Cocktail B in Micro-Tube (Red Cap)
- Cartridge containing Reagents 1, 2, 3, and 4
- Alexa Fluor 647 anti-mouse CD4 stain (AF647-CD4) [ordered separately]
- Alexa Fluor 647 anti-mouse CD8a stain (AF647-CD8) [ordered separately]

IsoCode Chip Set (-20°C)

Boxes of IsoCode Chips (2 per box)

IsoSpark: 4 chip kits

IsoLight: 4 or 8 chip kits

Cell Stain 405 [ordered separately]

Cell Stain 405 Diluent (DMSO) [part of cell stain 405 kit]

## Table 2: Required Consumables Not Supplied by Bruker

| Consumable | Type | Source | Catalog Number |
|---|---|---|---|
| T25 Flask | N/A | Corning | 353108 |
| T75 Flask | N/A | Corning | 430641U |
| 6 Well Plate Flat Bottom | N/A | Corning | 353046 |
| 96 Well Plate Flat Bottom | N/A | Corning | 353072 |
| MACS LS Column | N/A | Miltenyi | 130-042-401 |
| Centrifuge Tubes* | Polypropylene, 15 mL | VWR | CA62406-200 |
| Pipette Tips (Filtered) | 10 µL Graduated Filter Tips<br>100 µL Graduated Filter Tips<br>1000 µL XL Graduated Filter Tips | USA Scientific | 1181-3710<br>1183-1740<br>1182-1730 |
| Serological Pipette | 2 mL Pipette<br>5 mL Pipette<br>10 mL Pipette | USA Scientific | 1072-0510<br>1075-0110<br>1071-0810 |
| Lo-Bind Microcentrifuge Tubes, Sterile | 1.5 mL | USA Scientific | 4043-1081 |
| Syringe with BD Luer-Lok Tip | 10 mL | VWR | 75846-756 |
| 0.2 µm Syringe Filter with Acrylic Housing | N/A | VWR | 28145-501 |
| Fisherbrand Disposable PES Filter Units (0.20 µm) | 500 mL | Fisher Scientific | FB12566504 |

*Bruker strongly recommends that low protein binding centrifuge tubes are used for cell culture work to ensure optimal cell pelleting.

## Table 3: Required* Reagents Not Supplied by Bruker

| Reagent | Stock Concentration | Source | Catalog Number |
|---|---|---|---|
| RPMI | 1x | Fisher | MT10040CV |
| Penicillin-Streptomycin-Neomycin Solution Stabilized | 100x | Sigma | P4083-100mL |
| Glutamax | 100x | Thermo | 35050061 |
| FBS | 1x | Sigma | F2442-6X500mL |
| 2-Mercaptoethanol | 55 mM | Gibco | 21985-023 50mL |
| Recombinant IL-2 | 200 µg/mL | Biolegend | 589104 |
| Bovine Serum Albumin (BSA), lyophilized powder | N/A | Sigma-Aldrich | A9647-10G |
| Phosphate Buffered Saline (1XPBS) without Calcium or Magnesium | 1x | Gibco | 10010072 |
| RoboSep Buffer | 1x | StemCell Tech | 20104 |
| Ficoll Paque PREMIUM 1.084 | N/A | Cytiva | 17544602 |
| Miltenyi CD8a (Ly-2) Microbeads, Mouse, 2 mL | N/A | Miltenyi | 130-117-044 |
| Miltenyi CD4 (L3T4) Microbeads, Mouse, 2 mL | N/A | Miltenyi | 130-117-043 |
| Trypan Blue | 0.4% | Gibco | 15250-061 |
| CD3e Monoclonal Antibody (145-2C11), Functional Grade | 1 mg/mL | ThermoFisher/Invitrogen | 16-0031-86 |
| CD28 Monoclonal Antibody (37.51), Functional Grade | 1 mg/mL | ThermoFisher/Invitrogen | 16-0281-85 |
| Reagent alcohol 70% | N/A | Lab Grade | N/A |

*Reagents have been validated by Bruker and no alternatives may be used.

## Table 4: Cell Staining Reagents

| Test Material | Catalog number | Color |
|---|---|---|
| Cell Stain 405 | STAIN-1001-1 | Violet |
| Alexa Fluor 647 anti-mouse CD4 (AF647-CD4) | STAIN-1006-1 | Red |
| Alexa Fluor 647 anti-mouse CD8a (AF647-CD8) | STAIN-1007-1 | Red |

## Table 5: Required Equipment

| Equipment | Source | Catalog Number/Requirements |
|---|---|---|
| IsoLight, IsoSpark, or IsoSpark Duo Instrument | Bruker | ISOLIGHT-1000-1, ISOSPARK-1000-1, or ISOSPARK-1001-1 |
| Culture Hemocytometer | (Fisher) Hauser Levy | 02-671-55A |
| Hemocytometer Cover Glass | (Fisher) Hauser Levy | 02-671-53 |
| MidiMACS Separator | Miltenyi | 130-042-302 |
| MACS MultiStand | Miltenyi | 130-042-303 |

## Table 6: General Equipment

| Equipment | Requirements |
|---|---|
| Pipette | P10, P100, P200, P1000 |
| Pipettor | Ability to pipette between 1 and 10 mL |
| Incubator | 37°C, 5% $CO_2$ |
| Tabletop Centrifuge | Temperature controlled*; swinging bucket rotor; ability to centrifuge 15mL conical tubes |
| Microcentrifuge | Temperature controlled*; fixed rotor; ability to centrifuge 1.5 mL microcentrifuge tubes |
| Mini centrifuge | Ability to spin micro sample sizes |
| Water Bath | Ability to heat to 37°C |
| Microscope | Inverted light microscope with 10x and 20x objectives |
| Vortex Mixer | Ability to vortex vials and microcentrifuge tubes; adjustable speed |

*Temperature controlled centrifuges are required so that centrifuging steps can be conducted at room temperature without risk of overheating. Temperature on centrifuges should be set to 21°C.

# B. Before Getting Started

## 1. Important Precautions

Read MSDS documents of all materials prior to use.

### Working with Biohazardous Reagents

Please refer to your institute's guidelines and obtain proper training to handle potentially biohazardous samples. It is also strongly recommended that any lab personnel handling human samples should be vaccinated against HBV if the individual does not have sufficient HBV antibody titer.

Additional precautions need to be taken when working with samples that potentially contain an EID agent:

1. Laboratory workers should wear standard PPE, including disposable gloves, protective eyewear, and laboratory coats.
2. Any procedure or process that cannot be conducted in the designated EID BSC should be performed while wearing gloves, gown, goggles and a fit tested N-95 mask.
3. Work surfaces should be decontaminated on completion of work with appropriate disinfectants. This includes any surface that potentially comes in contact with the specimen (centrifuge, microscope, etc.).
4. All liquid waste produced in the processes must be treated to a final concentration of 10% bleach prior to disposal.

## 2. Reagents to Be Prepared Before Starting

### Table 7: Complete RPMI Recipe

- CRITICAL: Complete RPMI media has been validated for use by Bruker. Using alternative media may result in failed runs. Please contact your Field Application Scientist for additional information.

| Ingredient | Stock Concentration | Final Concentration | Amount for 500 mL | Vendor/Catalog |
|---|---|---|---|---|
| Penicillin-Streptomycin-Neomycin Solution Stabilized | 100x | 1x | 5 mL | Sigma/P4083-100mL |
| Glutamax | 100x | 1x | 5 mL | Thermo/35050061 |
| FBS | 100% | 10% | 50 mL | Sigma/F2442-6X500 mL |
| RPMI | 1x | 1x | 440 mL | Fisher/MT10040CV |

- CRITICAL: 2-Mercaptoethanol will be added to media used for all steps except cell staining quench media and final cell suspension volume to be loaded on IsoCode chip. 2-Mercaptoethanol should only be added to media right before use.

Note | Sterile-filter through 0.20 µm filter before use. Store complete RPMI Media at 4°C and warm up to 37°C in water bath prior to use.

## Table 8: 1% BSA Recipe

| Ingredient | Stock Concentration | Final Concentration | Amount for 100 mL | Vendor/Catalog |
|---|---|---|---|---|
| Bovine Serum Albumin (BSA), lyophilized powder | N/A | 1% | 1 g | Sigma-Aldrich/ A9647-10G |
| Phosphate Buffered Saline (1XPBS) without Calcium or Magnesium | 1X | 1X | 99 mL initially* | Gibco/10010072 |

*Rotate solution until BSA powder is dissolved and then bring final volume up to 100 mL with 1X PBS.

## Table 9: Working Stock of Recombinant IL-2 (1 µg/mL) Recipe

| Ingredient | Stock Concentration | Final Concentration | Amount for 10 mL | Vendor/Catalog |
|---|---|---|---|---|
| 1% BSA in 1X PBS (sterile filtered) | 1% | 1% | 9.95 mL | Table 8 |
| Recombinant IL-2 | 200 µg/mL | 1 µg/mL | 50 µL | BioLegend/589104 |

Note | Sterile-filter through a 0.2 µm PES filter before use.

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- CRITICAL: Prepare 200 µL IL-2 aliquots and freeze at -20°C for no longer than 1 month. Aliquots are single use only and are to be thawed immediately prior to their usage. If there is any remaining volume in an aliquot, do not refreeze but discard.

# C. Protocol

## Chapter 1: Getting Started

### Kit Contents

IsoLight IsoCode Reagent Box (4°C)

- 15 mL Tube A
- 15 mL Tube B
- 1.5 mL Tube A/B: Cocktail A in Micro-Tube (Green Cap) and Cocktail B in Micro-Tube (Red Cap)
- 50 mL Tubes Containing Reagents 1, 2, 3, 4, 5, 6, 7, 8
- 1 Bag of Disposable Reagent Sippers
- Alexa Fluor 647 anti-mouse CD4 stain (AF647-CD4) [ordered separately]
- Alexa Fluor 647 anti-mouse CD8a stain (AF647-CD8) [ordered separately]

IsoSpark IsoCode Reagent Box (4°C)

- 15 mL Tube A
- 15 mL Tube B
- 1.5 mL Tubes A/B: Cocktail A in Micro-Tube (Green Cap) and Cocktail B in Micro-Tube (Red Cap)
- Cartridge containing Reagents 1, 2, 3, and 4
- Alexa Fluor 647 anti-mouse CD4 stain (AF647-CD4) [ordered separately]
- Alexa Fluor 647 anti-mouse CD8a stain (AF647-CD8) [ordered separately]

IsoCode Chip Set (-20°C)

- Boxes of IsoCode Chips (2 per box)
  - IsoSpark: 4 chip kits
  - IsoLight: 4 or 8 chip kits
- Cell Stain 405 [ordered separately]
- Cell Stain 405 Diluent (DMSO) [part of cell stain 405 kit]

## Chapter 2: Coat Culture Plates with anti-CD3e

### Materials Required

| |
|---|
| Sterile 1X PBS (Room Temperature) |
| Anti-CD3e (Clone: 145-2C11) (4°C) 10 µg/mL |
| 15 mL Centrifuge Tube (*anti-CD3e Dilution*) |
| 96 Well Plate Flat Bottom (*Mouse T cell, Date, Time*) |

*All the following steps should take place in a sterile tissue culture hood.*

### Methods

- 1. Vortex the anti-CD3e tube at a slow speed for 10 seconds. TIP: Ensure contents are well suspended.
- 2. Spin anti-CD3e in mini centrifuge for 10 seconds. TIP: Ensure that contents are all in the bottom of the vial.

3. Preparation of anti-CD3e dilution in 1X PBS to a final concentration of 10 µg/mL in a 15 mL centrifuge tube:
    a. Dilute 20 µL of 1 mg/mL anti-CD3e into 2 mL of 1X PBS.
    b. Use 1 mL pipette to mix.
4. Pipette 100 µL of anti-CD3e dilution into 20 wells on a 96 well flat bottom plate. Wells coated with anti-CD3e dilution will be used for stimulation in Chapter 7. Leave the remaining wells empty for the unstimulated condition. NOTE: It is recommended to coat at least 2 wells per sample being run. Scale up the amount of anti-CD3e dilution if more than 20 coated wells are required. Note which wells have been coated.

- CRITICAL: Be careful not to create air bubbles.

5. Incubate covered plate at 4°C overnight.

## Chapter 3: Recovery of Cryopreserved Cells

### Materials Required

| |
|---|
| Complete RPMI (37°C) |
| 2-Mercaptoethanol (4°C) |
| Recombinant IL-2 at 1 µg/mL (-20°C) |
| Cryopreserved Mouse Cells |
| 15 mL Centrifuge Tube |
| Plate and/or Flask |
| For > 10 M cells, T75 Flask |
| For 6 - 9.9 M cells, T25 Flask |
| For < 6 M, 6 Well Plate |

*All the following steps should take place in a sterile tissue culture hood. Centrifugation steps should be performed at 21°C.*

### Methods

1. Prepare complete RPMI supplemented with 55 µM of 2-Mercaptoethanol.
    a. Dilute 50 µL of 55 mM 2-Mercaptoethanol into 50 mL of complete RPMI.
    b. Use serological pipette to mix thoroughly.

- CRITICAL: 2-Mercaptoethanol should only be added to media right before use. Discard any unused media.

2. Pipette 5 mL of complete RPMI/2-Mercaptoethanol into a 15 mL centrifuge tube, labeled *Thawed Mouse Cells*.

- 3. Using proper PPE, remove cells from liquid nitrogen storage and thaw cells. TIP: Be careful of contamination.

4. Quickly move vials into a water bath (37°C) to thaw. While thawing, swirl the vial in the water until a single ice crystal remains in the vial. Be sure to prevent (to the best of your ability) any of the water from the water bath from getting under the cap and into the sample.
5. When the sample is nearly thawed, remove the vial and immediately spray vial with 70% alcohol before bringing into the hood. It is important to allow the alcohol to evaporate before opening the vial.
- 6. Slowly pipette thawed cells into 5 mL of complete RPMI/2-Mercaptoethanol in 15 mL centrifuge tube, labeled *Thawed Mouse Cells*. TIP: Insert tip into complete RPMI when pipetting, be careful to not create bubbles.
- 7. Take 1 mL of complete RPMI/2-Mercaptoethanol and pipette into original thawed cell vial. Rinse inside the vial with the complete RPMI/2-Mercaptoethanol to recover additional thawed cells. TIP: Insert tip into complete RPMI, be careful to not create bubbles.
- 8. Draw up cell/complete RPMI/2-Mercaptoethanol mixture and pipette into the 15 mL centrifuge tube, labeled *Thawed Mouse Cells*. TIP: Insert tip into complete RPMI and pipette gently up and down. Be careful to not create bubbles.
9. Centrifuge cells for 10 minutes at 300 rcf.
10. While the cells are centrifuging, take the IL-2 (1 µg/mL) out from -20°C and thaw at room temperature.
- CRITICAL: Use IL-2 aliquot that has been frozen at -20°C for less than a month. Do not use IL-2 that has been previously thawed.
11. After cells are centrifuged, check for cell pellet.
- 12. Aspirate supernatant. TIP: Be careful not to aspirate cell pellet.
    a. Use pipette to remove last bit of supernatant.
13. Resuspend cell pellet in 1 mL of fresh complete RPMI/2-Mercaptoethanol.
- a. Mix well to resuspend. TIP: Make sure to pipette around the tube to ensure there are no clumps or bubbles.
14. Slowly add additional complete RPMI/2-Mercaptoethanol to a final concentration of 1 x $10^6$ cells/mL.
15. Mix thawed IL-2 thoroughly by carefully pipetting up and down.
16. Dilute 100 µL of 1 µg/mL IL-2 per 10 mL of cell suspension to a final concentration of 10 ng/mL IL-2.
- CRITICAL: Discard thawed IL-2 aliquot if there is any volume remaining. IL-2 must only be thawed once.
- 17. Mix with serological pipette. TIP: Gently pipet up and down 3-5 times, be careful to not create bubbles.
- 18. Transfer cell suspension to appropriate flask or plate. TIP: Slowly pipette down the side of the flask as to not create bubbles.
- 19. Spread out cell suspension by rocking the flask or plate carefully to fully cover the bottom of the container. TIP: Be careful to not make bubbles.
20. Move to incubator for overnight recovery at 37°C, 5% $CO_2$. NOTE: The time period for overnight recovery is considered 16 – 20 hours, but not exceeding 24 hours.

# Chapter 4: Post-Recovery Sample Setup

## Materials Required

| |
|---|
| Complete RPMI (37°C) |
| 2-Mercaptoethanol (4°C) |
| 15 mL Centrifuge Tube |
| Overnight Recovered Cells from Chapter 3 or Fresh Mouse Cells if Working with Fresh Samples |
| Lo-Bind Microcentrifuge Tube for Cell Count |

*All the following steps should take place in a sterile tissue culture hood. Centrifugation steps should be performed at 21°C.*

## Methods

1. Prepare complete RPMI supplemented with 55 µM of 2-Mercaptoethanol.
    a. Dilute 50 µL of 55 mM 2-Mercaptoethanol into 50 mL of complete RPMI.
    b. Use serological pipette to mix thoroughly.

● CRITICAL: 2-Mercaptoethanol should only be added to media right before use. Discard any unused media.

2. Transfer cells from flask or plate into 15 mL centrifuge tube.

● 3. Add complete RPMI/2-Mercaptoethanol to flask or plate and rinse 5 times. TIP: Make sure to spread out the complete RPMI to gather maximum number of cells.
    - For T75 Flask add 3 mL
    - For T25 Flask add 2 mL
    - For 6 Well Plate add 1 mL

4. Transfer cell/complete RPMI mixture/2-Mercaptoethaol to the 15 mL centrifuge tube.

● 5. Mix well 5 times with 10 mL serological pipette. TIP: Be careful not to create bubbles.

● 6. Take a 10 µL aliquot of your cells and transfer to a Lo-Bind Microcentrifuge Tube for cell counting. CRITICAL: See Appendix D1 for cell counting instructions.

● 7. Centrifuge cells for 10 minutes at 300 rcf. While cells are centrifuging, use hemocytometer to count cells. CRITICAL: See Appendix D1 for cell counting instructions.

● CRITICAL: If cells are less than 80% viable, proceed to Appendix D2 Protocol: Dead Cell Removal Using Ficoll.

8. Proceed immediately to next chapter.



Key: ● TIP, ● CRITICAL, ● OPTIONAL

# Chapter 5: CD8 Sample Enrichment

## Materials Required

Complete RPMI (37°C)
2-Mercaptoethanol (4°C)
RoboSep Buffer (4°C)
Miltenyi CD8a (Ly-2) Microbeads, Mouse, 2 mL (4°C)
MACS LS Column
Prepared Cells from Chapter 4
Enrichment Kit:
- MACS Metal Plate/Magnet Kit
- 3 x 15 mL Centrifuge Tubes (*Discard, Flow Through, CD8 fraction*)
- Lo-Bind Microcentrifuge Tube for Post-Enrichment CD8

*All the following steps should take place in a sterile tissue culture hood. Centrifugation steps should be performed at 21°C.*

## Methods

1. Remove the centrifuged cells and check for cell pellet.
2. Aspirate supernatant. TIP: Be careful not to aspirate the cells.
    a. Use pipette to aspirate remaining supernatant.

CRITICAL: For every 1 x $10^7$ cells, resuspend in 90 µL RoboSep (4°C) and 10 µL of CD8a beads (4°C).

3. Add 90 µL of cold RoboSep to 15 mL centrifuge tube containing 1 X $10^7$ or fewer cells.
4. Vortex the Miltenyi CD8a (Ly-2) Microbeads at a slow speed for 10 seconds.
5. Add 10 µL of Miltenyi CD8a (Ly-2) Microbeads to the cell suspension in RoboSep (containing 1 X $10^7$ or fewer cells) and mix well by gently pipetting up and down 5 times.

TIP: Make sure to eliminate clumps so that beads are evenly distributed among cells. Be careful not to create bubbles.

6. Incubate in refrigerator (4°C) for 10 minutes. TIP: Don't incubate on ice as increased incubation times may be required.
7. Set up MACS sorting by setting metal plate in tissue culture hood and placing magnet on metal plate. Place LS column in magnet with wings facing out and align the 15 mL centrifuge tube labeled "Discard" under the LS column. CRITICAL: LS Column should not touch the tubes.
8. After incubating for 10 minutes, for 1 x $10^7$ cells or fewer, add 400 µL of cold RoboSep to the 15 mL tube to bring up volume to 500 µL.
    a. Mix well to resuspend by gently pipetting up and down 5 times. TIP: Make sure to pipette around the tube to ensure there are no clumps or bubbles.

TIP: Keep RoboSep in refrigerator during enrichment process.

CRITICAL: Be careful not to let column dry out. Do not add liquid when there is already liquid in the LS Column.



Key: ● TIP, ● CRITICAL, ● OPTIONAL

9. Start with the LS column over the "Discard" tube, add 3 mL of cold RoboSep to LS Column. CRITICAL: Be careful not to create bubbles or touch sides of LS column. Let all the RoboSep flow through before moving on to the next step. As a reminder, be careful to not let the column dry out.
10. Unscrew and keep cap for "Flow Through" tube. NOTE: This is in preparation for next step to ensure the column does not dry out during the transition.
11. When the last drop falls through to the "Discard" tube, move the rack over so the LS column is over the "Flow Through" tube. CRITICAL: Be careful not to let column dry. If there is one drop remaining that will not fall, move on to the next step.
12. Increase volume of pipette to 800 µL to ensure all 500 µL of the cell suspension is drawn up.
13. Mix cell suspension by gently pipetting up and down 5 times. NOTE: This ensures that the cells are evenly dispersed after sitting.
14. Draw up all 500 µL of cell suspension and pipette carefully into the center of the LS column without touching sides of the column.
15. Wash LS column with 3 mL of cold RoboSep.
    a. Rinse inside walls of cell suspension tube with 3 mL of cold RoboSep before transferring the mixture to LS Column. NOTE: This is to retrieve any cells that have been left behind.
    i. Pipette all the mixture into LS Column after last drop passes through or does not fall from step 14. CRITICAL: Be careful not to let LS Column dry out or allow pipette to touch sides.
16. After the last drop of the wash passes through or does not fall, remove the LS Column carefully from the magnet, and place carefully on the tube labeled for "CD8 fraction."
17. Cap the "Flow Through" tube, this will be used for the CD4 enrichment. Do not discard.
18. Add 5 mL of cold RoboSep to the LS column. CRITICAL: Be careful not to touch the sides.
19. Take plunger, smoothly push down on the plunger to push the RoboSep buffer through the LS Column. TIP: Lift up at the end of the plunging action so that the liquid does not splash back onto the LS Column tip.
20. Set LS Column back on the "CD8 fraction" tube.

CRITICAL: Do not allow the plunger to interact with external contaminants. It will be used for one more step.

21. Loosen up plunger. Remove plunger briefly from column and hold in one hand.
22. Add another 2 mL of cold RoboSep to the LS Column.
23. Take plunger, smoothly push down on the plunger to push the RoboSep buffer through the LS Column. TIP: Lift up at the end of the plunging action so that the liquid does not splash back onto the LS Column tip.
24. Discard LS Column and plunger.
25. Centrifuge "CD8 fraction" tube and "Flow Through" (CD8 Depleted Cell Fraction) tube for 10 minutes at 300 rcf.
26. Prepare complete RPMI supplemented with 55 µM of 2-Mercaptoethanol.
    a. Dilute 5 µL of 55 mM 2-Mercaptoethanol into 5 mL of complete RPMI.

b. Use serological pipette to mix thoroughly.

- CRITICAL: 2-Mercaptoethanol should only be added to media right before use. Discard any unused media.

27. After cells are centrifuged, check for cell pellets.

- 28. Aspirate RoboSep buffer from "CD8 fraction" and "Flow Through" (CD8 Depleted Cell Fraction) tubes. TIP: Be careful to not aspirate cell pellet.

29. Use a pipette to aspirate the remaining supernatant from each tube. TIP: Be careful to not aspirate cell pellet.

- 30. Add 1 mL complete RPMI/2-Mercaptoethanol to "CD8 fraction" and resuspend cell pellet. TIP: Make sure there are no clumps or bubbles.

31. Add an additional 1 mL of complete RPMI/2-Mercaptoethanol and mix thoroughly by gently pipetting up and down 5 times. TIP: Make sure there are no clumps or bubbles.

- 32. Aliquot 10 µL of the "CD8 fraction" into a Lo-Bind Microcentrifuge tube and proceed to cell count. CRITICAL: See Appendix D1 for cell counting instructions.

33. Move "CD8 fraction" tube to incubator until Cell Stimulation (Chapter 7).

## Chapter 6: CD4 Sample Enrichment

### Materials Required

Complete RPMI (37°C)
2-Mercaptoethanol (4°C)
RoboSep Buffer (4°C)
Miltenyi CD4 (L3T4) Microbeads, Mouse, 2 mL (4°C)
MACS LS Column
CD8 Depleted Fraction or Overnight Recovered Cells from Chapter 4 if CD8 Depletion is Not Necessary
Enrichment Kit:
MACS Metal Plate/Magnet Kit
3 x 15 mL Centrifuge Tubes (*Discard, Flow Through, CD4 fraction*)
Lo-Bind Microcentrifuge Tube for Post-Enrichment CD4

*All the following steps should take place in a sterile tissue culture hood. Centrifugation steps should be performed at 21°C.*

### Methods

- CRITICAL: For every 1 x $10^7$ cells, resuspend in 90 µL RoboSep (4°C) and 10µL of CD4 (L3T4) beads (4°C).

1. Add 90 µL of cold RoboSep to 15 mL centrifuge tube containing CD8 depleted fraction of 1 X $10^7$ or fewer cells.
2. Vortex the Miltenyi CD4 (L3T4) Microbeads at a slow speed for 10 seconds.
3. Add 10 µL of Miltenyi CD4 (L3T4) Microbeads to the cell suspension in RoboSep (containing 1 X $10^7$ or fewer cells) and mix well by gently pipetting up and down 5 times.

- TIP: Make sure to eliminate clumps so that beads are evenly distributed among cells. Be careful not to create bubbles.

4. Incubate in refrigerator (4°C) for 10 minutes. TIP: Don't incubate on ice as increased incubation times may be required.

5. Set up MACS sorting by setting metal plate in tissue culture hood and placing magnet on metal plate. Place LS column in magnet with wings facing out and align the 15 mL centrifuge tube labeled "Discard" under the LS column. CRITICAL: LS Column should not touch the tubes.

6. After 10 minutes, for 1 x $10^7$ cells or fewer, add 400 µL of cold RoboSep to the 15 mL tube to bring up volume to 500 µL.

    a. Mix well to resuspend by gently pipetting up and down 5 times. TIP: Make sure to pipette around the tube to ensure there are no clumps or bubbles.

- TIP: Keep RoboSep in refrigerator during enrichment process.
- CRITICAL: Be careful not to let column dry out. Do not add liquid when there is already liquid in the LS Column.

7. Start with the LS column over the "Discard" tube, add 3 mL of cold RoboSep to LS Column. CRITICAL: Be careful not to create bubbles or touch sides of LS column. Let all the RoboSep flow through before moving on to next step. As a reminder, be careful to not let the column dry out.

8. Unscrew and keep cap for "Flow Through" tube. NOTE: This is in preparation for next step to ensure the column does not dry out during the transition.

9. When last drop falls through to "Discard" tube, move the rack over so the LS column is over the "Flow Through" tube. CRITICAL: Be careful not to let column dry. If there is one drop remaining that will not fall, move on to the next step.

10. Increase volume of pipette to 800 µL to ensure all 500 µL of the cell suspension is drawn up.

11. Mix cell suspension by gently pipetting up and down 5 times. NOTE: This ensures that the cells are evenly dispersed after sitting.

12. Draw up all 500 µL of cell suspension and pipette carefully into the center of the LS column without touching sides of the column.

13. Wash LS column with 3 mL of cold RoboSep.

    a. Rinse inside walls of cell suspension tube with 3 mL of RoboSep before transferring the mixture to LS Column. NOTE: This is to retrieve any cells that have been left behind.

        i. Pipette all of the mixture into LS Column after last drop passes through or does not fall from step 12. CRITICAL: Be careful not to let LS Column dry out or allow pipette to touch sides.

14. After the last drop of the wash passes through or does not fall, remove the LS Column carefully from the magnet, and place carefully on the tube labeled for "CD4 Fraction."

15. Cap the "Flow Through" tube and discard.

16. Add 5 mL of cold RoboSep to the LS column. CRITICAL: Be careful not to touch the sides.



Key: ● TIP, ● CRITICAL, ● OPTIONAL

- 17. Take plunger, smoothly push down on the plunger to push the RoboSep buffer through the LS Column. TIP: Lift up at the end of the plunging action so that the liquid does not splash back onto the LS Column tip.
18. Set LS Column back on the "CD4 Fraction" tube.
19. Loosen up plunger. Remove plunger briefly from column and hold in one hand.
- CRITICAL: Do not allow the plunger to interact with external contaminants. It will be used for one more step.
20. Add another 2 mL of cold RoboSep to the LS Column.
- 21. Take plunger, smoothly push down on the plunger to push the RoboSep buffer through the LS Column. TIP: Lift up at the end of the plunging action so that the liquid does not splash back onto the LS Column tip.
22. Discard LS Column and plunger.
23. Centrifuge "CD4 Fraction" tube for 10 minutes at 300 rcf.
24. Prepare complete RPMI supplemented with 55 µM of 2-Mercaptoethanol.
    a. Dilute 5 µL of 55 mM 2-Mercaptoethanol into 5 mL of complete RPMI.
    b. Use serological pipette to mix thoroughly.
- CRITICAL: 2-Mercaptoethanol should only be added to media right before use. Discard any unused media.
25. After cells are centrifuged, check for cell pellet.
- 26. Aspirate RoboSep buffer from "CD4 fraction" tube. TIP: Be careful to not aspirate cell pellet.
- 27. Use pipette to aspirate the remaining supernatant from each tube. TIP: Be careful to not aspirate cell pellet.
- 28. Add 1 mL RPMI/2-Mercaptoethanol to "CD4 fraction" and resuspend cell pellet. TIP: Make sure there are no clumps or bubbles.
- 29. Add an additional 1 mL of RPMI/2-Mercaptoethanol and mix thoroughly by gently pipetting up and down 5 times. TIP: Make sure there are no clumps or bubbles.
- 30. Aliquot 10 µL of the CD4 fraction into a Lo-Bind Microcentrifuge tube and proceed to cell count. CRITICAL: See Appendix D1 for cell counting instructions.
31. Move "CD4 fraction" tube to incubator until Cell Stimulation. (Chapter 7).

## Chapter 7: Cell Stimulation

Materials Required

| |
|---|
| Complete RPMI (37°C) |
| 2-Mercaptoethanol (4°C) |
| Anti-CD28 (4°C) |
| Sterile 1X PBS (Room Temperature) |
| Prepared anti-CD3e Coated 96 Well Plate from Chapter 2 |
| 15 mL Centrifuge Tubes |
| Incubated CD8 Tube in complete RPMI |
| Incubated CD4 Tube in complete RPMI |

*All the following steps should take place in a sterile tissue culture hood. Centrifugation steps should be performed at 21°C.*

Methods

1. Prepare complete RPMI supplemented with 55 µM of 2-Mercaptoethanol.
    a. Dilute 10 µL of 55 mM 2-Mercaptoethanol into 10 mL of complete RPMI.
    b. Use serological pipette to mix thoroughly.

- CRITICAL: 2-Mercaptoethanol should only be added to media right before use. Discard any unused media.

2. Prepare Complete RPMI/2-Mercaptoethanol Unstimulated media.
    a. Aliquot 5 mL complete RPMI/2-Mercaptoethanol into a 15 mL Centrifuge Tube labeled "Complete RPMI Unstimulated" —set this complete RPMI/2-Mercaptoethanol aside as it will serve as the media used for the **unstimulated** (negative control) condition.

- CRITICAL: Do not add stimulants into this complete RPMI/2-Mercaptoethanol. Volume required is dependent on number of cells.

- 3. Vortex anti-CD28 at a slow speed for 10 seconds. TIP: Ensure contents are well suspended.
- 4. Spin anti-CD28 in a mini centrifuge for 10 seconds. TIP: Ensure that contents are all in the bottom of the vial.
- 5. Prepare anti-CD28 and complete RPMI/2-Mercaptoethanol mixture by supplementing complete RPMI with 5 µg/mL of anti-CD28 into a 15 mL centrifuge tube labeled "CD28 Complete RPMI". CRITICAL: Volume is dependent on number of cells.
    a. Dilute 25 µL of 1 mg/mL anti-CD28 into 5 mL of complete RPMI/2-Mercaptoethanol.
    b. Use serological pipette to mix thoroughly.
6. Take CD8 and CD4 cells from incubator.
7. Centrifuge CD8 and CD4 cells for 10 minutes at 300 rcf.
8. After cells are centrifuged, check for cell pellet.
- 9. Aspirate supernatant with pipette. TIP: Make sure to use a manual pipette to prevent accidental aspiration of cell pellet.
- 10. Using the complete RPMI/2-Mercaptoethanol set aside for the **unstimulated** condition, resuspend all the CD8 and CD4 cells with complete RPMI/2-Mercaptoethanol to a cell concentration of 1 x $10^6$ cells/mL. TIP:

This step is for preparing the unstimulated (negative control) cells. This complete RPMI/2-Mercaptoethanol is not supplemented with any stimulants.

- 11. Remove previously prepared anti-CD3e-coated 96 well plate from refrigerator.
- 12. Plate 100 µL of the CD8 **unstimulated** cells, per uncoated and empty well, on the 96 well flat bottom plate. TIP: Plate at least two wells on the 96 well flat bottom plate for the unstimulated condition.
13. Repeat step using CD4 cells.
14. Centrifuge the remaining CD4 and CD8 cells (that will serve as the CD3/CD28 **stimulated** cells) for 10 minutes at 300 rcf.
15. While cells are centrifuging, aspirate the anti-CD3e dilution from 96 well plate. TIP: Aspirate from the edge of well.
16. Add 100 µL of 1X PBS to each well previously coated with anti-CD3e to rinse.
17. After cells are centrifuged, check for cell pellet.
- 18. Aspirate supernatant with pipette. TIP: Make sure to use a manual pipette to prevent accidental aspiration of cell pellet.
19. Use a serological pipette to mix the anti-CD28 and complete RPMI/2-Mercaptoethanol mixture to ensure it is evenly distributed.
- 20. Using the anti-CD28/complete RPMI/2-Mercaptoethanol mixture from step 19, resuspend CD8 cells to a cell concentration of 1 x $10^6$ cells/mL. CRITICAL: When resuspending, take into consideration the cells removed for the unstimulated condition. TIP: Resuspend as thoroughly as possible, but gently.
21. Repeat step using CD4 cells.
- 22. Aspirate 1X PBS from each well previously coated with anti-CD3e. TIP: Aspirate from the edge of the well.
- 23. Mix CD8 cells by pipetting up and down gently 5 times. Add 100 µL of cell suspension to **coated wells** on plate. NOTE: It is suggested to use a maximum of half the coated wells for CD8 cells and leave the remaining coated wells for CD4 cells. CRITICAL: Be careful not to create bubbles. This will maximize even stimulation of cell suspension.
24. Repeat step using CD4 cells.
25. Incubate plate for 48 hours at 37°C, 5% $CO_2$.

## Chapter 8: Chip Thaw

Materials Required

IsoCode Chips in Vacuum Sealed Bag (-20°C)

Methods

1. Take vacuum sealed bag containing IsoCode chips from -20°C. CRITICAL: Chips must stay sealed until Chip Loading (Chapter 10).
2. Place on bench to thaw at ambient temperature 30 - 60 minutes prior to use.
3. While chips thaw, prepare liquid reagents and setup in the Bruker instrument. Refer your instrument's system guide for detailed instructions.

## Chapter 9: Cell Staining

NOTE: Please read before proceeding with cell staining.

There are 2 options for cell staining:

1. Cell staining (violet) as described in Chapter 9a.
2. Dual staining with cell stain 405 (violet) and either CD4 or CD8 specific surface marker stains (red). This is achieved by following Chapter 9a and Chapter 9b sequentially.

Use of CD8 and/or CD4 specific surface marker staining is recommended for TILs or T cells in a debris rich environment where enrichment alone does not guarantee a high purity sample for loading onto the IsoCode chip.

### Cell Staining Reagents

| Test Material | Catalog Number | Color |
|---|---|---|
| Cell Stain 405 | STAIN-1001-1 | Violet |
| Alexa Fluor 647 anti-mouse CD4 (AF647-CD4) | STAIN-1006-1 | Red |
| Alexa Fluor 647 anti-mouse CD8a (AF647-CD8) | STAIN-1007-1 | Red |

## Chapter 9a: Cell Staining (Violet)

Materials Required

Anti-CD3e/CD28 Stimulated & Unstimulated CD8 & CD4 Cells in 96 Well Plate from Chapter 7
Lo-Bind Microcentrifuge Tubes (*Stain Master Mix, CD8, CD4*)
Sterile 1X PBS (Room Temperature)
Complete RPMI (37°C)
Cell Stain 405 (-20°C)
Cell Stain 405 Diluent (DMSO) (-20°C)

*All the following steps should take place in a sterile tissue culture hood. Centrifugation steps should be performed at 21°C.*

Methods

1. Prepare cell stain 405 stock.
    a. Thaw tube of cell stain 405 diluent (DMSO) at room temperature.
    b. Spin tubes of cell stain 405 and cell stain 405 diluent (DMSO) in a mini centrifuge for 10 seconds to collect the contents at the bottom of the tubes.
    c. Add 20 µL of cell stain 405 diluent (DMSO) directly to the tube of cell stain 405. Pipet up and down 15 times gently to resuspend.

- CRITICAL: Cell stain 405 must be prepared fresh. Discard remaining stain – do not store.

- 2. Prepare stain master mix by diluting 1 µL of cell stain 405 into 1 mL of 1X PBS in a Lo-Bind microcentrifuge tube (1:1000 final dilution). With the same pipette tip, pipette up and down 10 times to ensure all cell stain 405 has been released. Depending on sample number and cell count, additional tubes of stain master mix may need to be prepared. CRITICAL: Failure to follow these steps will negatively impact cell counts.
    a. With a P1000 set to 500 µL, gently pipette the stain master mix up and down 15 times.
    b. Gently vortex the stain master mix for 5 seconds.
    c. Ensure master mix is mixed well before adding stain to cells.
3. Remove 96 well plate with CD8 and CD4 cells from incubator.
4. Mix CD8 cells by pipetting up and down. Transfer cells to a Lo-Bind microcentrifuge tube by using P100 pipette to draw up 100 µL at a time in a gentle, circular motion until well is empty. NOTE: Pool wells if there are replicates.
5. Repeat step 4 using CD4 cells.
6. Centrifuge cells for 10 minutes at 300 rcf.
7. After cells are centrifuged, check for cell pellets.

- 8. Remove the majority of the supernatant with a P1000 or P200 pipette. Switch to a P10 pipette to remove all media from the cell pellet.* TIP: Be careful not to aspirate the cell pellets.

    *NOTE: Supernatants may be stored at -80°C for bulk assay.

9. Add 1 mL of PBS to dilute any remaining media and mix by pipetting up and down.

- CRITICAL: Failure to remove excess media will result in poor staining.

10. Centrifuge cells for 10 minutes at 300 rcf.
11. After cells are centrifuged, check for cell pellets.

- 12. Remove the majority of the supernatant with a P1000 or P200 pipette. Switch to a P10 pipette to remove all media from the cell pellet. TIP: Be careful not to aspirate the cell pellets.

13. Gently remix stain master mix.

- CRITICAL: Failure to remix stain master mix will result in poor staining.

- 14. For every 1 x $10^6$ cells, add 100 µL of well mixed stain master mix to each cell suspension tube. CRITICAL: Pipet to mix the cells 15 times. Be careful to not create bubbles.
15. Incubate for 5 minutes at 37°C in the dark.
- 16. Gently pipette to mix the cell suspension 15 times. CRITICAL: Be careful to not create bubbles.
17. Incubate for an additional 5 minutes at 37°C in the dark.
- 18. After incubation, add 5 times the volume of complete RPMI without 2-Mercaptoethanol. CRITICAL: Pipet to mix the cells 15 times. Be careful to not create bubbles.
19. Incubate for 10 minutes at 37°C in the dark.
- 20. Take 10 µL of cells to count. Count cells using a hemocytometer and determine percent of viable cells as described in Appendix D1. TIP: Cell counting can be done while cells are incubating.
21. Centrifuge stained cells for 10 minutes at 300 rcf.
22. After cells are centrifuged, check for cell pellets.
- 23. Aspirate supernatant with a pipette. TIP: Be careful not to aspirate the cell pellets.

    NOTE: If dual staining with anti-CD4 and anti-CD8a, proceed immediately to Chapter 9b.

24. If only staining with cell stain 405, resuspend cell pellet with complete RPMI without 2-Mercaptoethanol to a cell density of 1 x $10^6$ cells/mL and proceed to Chapter 10.

## Chapter 9b: Surface Marker Specific Staining (Optional)

| |
|---|
| Cells Stained with Cell Stain 405 from Chapter 9a |
| Complete RPMI (37°C) |
| AF647 anti-mouse CD4 (4°C) |
| AF647 anti-mouse CD8a (4°C) |
| Lo-Bind Microcentrifuge Tube for Cell Count |

*All the following steps should take place in a sterile tissue culture hood. Centrifugation steps should be performed at 21°C.*

Methods

- CRITICAL: For every 1 x $10^5$ cells, resuspend in 18 µL of complete RPMI and 2 µL of appropriate surface marker stain.

1. Resuspend the cell pellets with 18 µL complete RPMI without 2-Mercaptoethanol for 1 x $10^5$ or fewer cells.
2. Spin tubes of AF647 anti-mouse CD4 and CD8 in a mini centrifuge for 10 seconds to collect stain at the bottom of the tube.

- 3. Add 2 µL of AF647 anti-mouse CD4 to CD4+ samples (1:10 final dilution) for 1 x $10^5$ or fewer cells. Mix gently by pipetting up and down. CRITICAL: Be sure to use appropriate stain for each cell subset.
- 4. Add 2 µL of AF647 anti-mouse CD8a to CD8+ samples (1:10 final dilution) for 1 x $10^5$ or fewer cells. Mix gently by pipetting up and down. CRITICAL: Be sure to use appropriate stain for each cell subset.

5. Incubate for 20 minutes at room temperature in the dark.
- 6. After incubation, add 1 mL of complete RPMI without 2-Mercaptoethanol to each sample tube. TIP: Mix gently, be careful not to create bubbles.
- 7. Take 10 µL of cells to count. Count cells using a hemocytometer and determine percent of viable cells as described in Appendix D1. TIP: Cell counting can be done while cells are incubating.
8. Centrifuge stained cells for 10 minutes at 300 rcf.
9. After cells are centrifuged, check for cell pellets.
- 10. Aspirate supernatant with a pipette. TIP: Be careful not to aspirate the cell pellets.
- 11. Resuspend the cell pellet with complete RPMI without 2-Mercaptoethanol to a cell density of 1 x $10^6$ cells/mL. Proceed to Chapter 10.

## Chapter 10: Chip Loading

Materials Required (Pre-prepared)

| |
|---|
| Pre-Thawed IsoCode Chips in Vacuum Sealed Bag from Chapter 8 |
| Stained CD8 Cells at 1 x $10^6$ cells/mL |
| Stained CD4 Cells at 1 x $10^6$ cells/mL |

Methods

- 1. Remove IsoCode chips from vacuum sealed bag and place on a flat surface. CRITICAL: Keep protective blue film on bottom of chip.
- 2. Resuspend CD8 fractions by gently pipetting up and down 15 times. Immediately proceed to chip loading. Pipette 30 µL of cell suspension into IsoCode chip. CRITICAL: Be careful not to create bubbles. Insert pipette tip vertically into inlet port until tip lightly touches bottom, and slowly pipette 30 µL into inlet port. Be careful not to eject second step of pipette–it will cause bubbles.
- 3. Resuspend CD4 fractions by gently pipetting up and down 15 times. Immediately proceed to chip loading. Pipette 30 µL of cell suspension into IsoCode chip. CRITICAL: Be careful not to create bubbles. Insert pipette tip vertically into inlet port until tip lightly touches bottom, and slowly pipette 30 µL into inlet port. Be careful not to eject second step of pipette–it will cause bubbles.
4. Let IsoCode chips sit for one minute on a flat surface.
- 5. Check bottom of chip to ensure liquid has entered the chip. TIP: If liquid has not flowed, tap IsoCode chip on flat surface lightly.
6. When inserting IsoCode chip into the instrument, make sure the logo is facing up and towards you with the magnet facing the instrument. Take the blue film off while inserting each IsoCode chip into the instrument.

NOTE: Please refer your instrument's loading instructions for details.

## D: Appendix

## D1 Protocol: Cell Quantification & Viability

Materials Required

| |
|---|
| Hemocytometer |
| 10 µL aliquot of cells |
| Trypan Blue |

NOTE: Automated cell counters can be used in this protocol EXCEPT prior to loading cells on chip due to spectral overlap of the stains. Manual cell counting is required prior to loading on the chip.

NOTE: To obtain an accurate representation of cell viability, cells should be counted within 15 minutes of staining as cell viability will drop over time because Trypan Blue is toxic.

1. Quick spin the Trypan Blue to pellet potential debris. Remove aliquot from the top of Trypan Blue.
2. Using a P10 pipette, add equal volume of Trypan blue solution to 10 µL of sample. Mix gently to resuspend.
   TIP: Make sure to pipette around the tube to ensure there are no clumps or bubbles.
3. Load onto hemocytometer. CRITICAL: Be careful not to overfill or create bubbles.
4. Count and record viable (clear) and dead cells (blue) of all four 16-square corners.

- CRITICAL: If more than 200 cells/16 squares were counted, repeat count using a 1:5 or 1:10 dilution with 1X PBS or complete RPMI using a fresh sample aliquot.

5. Calculate the concentration of cells as follows:
   a. Concentration (cells/mL) = Average per square cell count x $10^4$ x dilution factor
6. Calculate the number of cells as follows:
   a. Number of cells = Cell concentration (cells/mL) from D.1.5 x total volume of cell suspension (mL)
7. Calculate percent viable cells:
   a. % Viable cells = 100 x number of viable cells / [number of viable cells + number of dead cells]

## D2 Protocol: Dead Cell Removal Using Ficoll

Materials Required

| |
|---|
| Complete RPMI (37°C) |
| 2-Mercaptoethanol (4°C) |
| Cells (Minimum $3x10^6$) |
| 2 x 15 mL Centrifuge Tubes |
| Lo-Bind Microcentrifuge Tubes |
| Ficoll Paque PREMIUM 1.084 |

- CRITICAL: It is recommended to start this protocol with a minimum of $3x10^6$ total cells.

1. Carefully add 6 mL of Ficoll to the bottom of the required number of 15 mL centrifuge tube(s) prior to harvesting stimulation cultures.
2. Centrifuge cells for 10 minutes at 300 rcf.

3. Remove cells from centrifuge, check for cell pellet.
- 4. Aspirate supernatant. TIP: Be careful not to aspirate cell pellet.
    a. Use pipette to aspirate remaining supernatant.
- 5. Resuspend the pellet(s) in 7 mL of complete RPMI. TIP: Be careful not to create bubbles.
- CRITICAL: Do not use more than 1 x $10^7$ cells of your suspension per Ficoll tube.
- 6. Add the cell suspension(s) VERY SLOWLY to the tube(s) containing Ficoll. CRITICAL: Place the tip of your pipette on the wall of the tube, close to the Ficoll layer. Add cell suspension VERY SLOWLY.
- CRITICAL: This step must be done carefully and slowly to avoid mixing of the layers.
7. Centrifuge tubes for 20 minutes at 300 rcf without brake or acceleration.
- CRITICAL: Turn acceleration and brakes off to preserve the density layers established during centrifugation.
8. Prepare complete RPMI supplemented with 55 µM of 2-Mercaptoethanol.
    a. Dilute 6 µL of 55 mM 2-Mercaptoethanol into 6 mL of complete RPMI.
    b. Use serological pipette to mix thoroughly.
- CRITICAL: 2-Mercaptoethanol should only be added to media right before use. Discard any unused media.
9. While cells centrifuge, prepare appropriate number of 15 mL centrifuge tube(s) containing 6 mL of complete RPMI/2-Mercaptoethanol.
10. Remove cells from centrifuge, check for cloudy layer which are the viable cells.
- 11. Aspirate a small volume of the supernatant. CRITICAL: Be careful not to aspirate cloudy layer containing viable cells.
12. Using a P1000 pipette, collect the viable cells by recovering the cloudy layer between Ficoll and complete RPMI media
13. Transfer cells into the 15 mL centrifuge tube(s) containing complete RPMI.
- 14. Aliquot 10 µL of cell/complete RPMI mixture(s) into a Lo-Bind Microcentrifuge Tube(s) and proceed to cell count. CRITICAL: See Appendix D1 for cell counting instructions.



Key: ● TIP, ● CRITICAL, ● OPTIONAL

## Troubleshooting & References

Please contact Support at 844-476-7539 (toll free) or 475-221-8402 or email support@isoplexis.com with specific troubleshooting questions.

| Problem | Possible Reason | Solution |
|---|---|---|
| Low quality cell count on chip *Cell Counting & Concentration related* | • Recommended cell concentrations not used<br>• Issue with Cell Counting procedure<br>• Trypan Blue may have debris<br>• Trypan Blue is toxic<br>• Poor cell removal from anti-CD3 plate | • Use recommended cell concentrations during incubation (**Chapter 7**)<br>• Use appropriate dilutions recommended in **Appendix D1**<br>• Do a recount if initial count does not seem accurate<br>• Quick spin Trypan Blue to pellet potential debris, remove aliquot from top of Trypan Blue. Start with fresh aliquot of Trypan Blue.<br>• Count within 15 minutes of staining the cells<br>• Thoroughly mix cells in well with pipette prior to transferring to tube (refer to **steps 9a.4 and 9a.5**) |
| Low quality cell count on chip *Stain Process related* | • Use of media other than the recommended media in protocol which could interact with cell stain<br>• Use of stains not recommended in protocol<br>• Recommended stain concentration, incubation time and/or incubation temperature not used<br>• Cell stain 405 was stored prior to use<br>• Media not completely removed from cell pellet prior to staining | • Use complete RPMI media following recipe in **Table 7**<br>• Use Bruker provided validated stain (**Table 4: Cell Staining Reagents**)<br>• Follow staining steps as highlighted in **Chapter 9**<br>• Use only freshly prepared cell stain 405 per **Chapter 9a**.<br>• Ensure all media is removed from cell pellet in **step 9a.8** |
| Low quality cell count on chip *Technique Detail related* | • Bubbles loaded onto chip, especially at Chip Loading<br>• Detection of potential artifacts such as debris, cell clumping, inefficient enrichment possibly due to:<br>  ○ Pipetting wrong concentration<br>  ○ Not fully inserting column into MACS separator<br>  ○ Reagents not stored at recommended temperatures<br>• Recommended number of cells not loaded on chip<br>• Cell pellet or cells lost during centrifuging | • Follow Critical step in **10.2** and **10.3** to avoid introduction of bubbles on chip<br>• Ensure use of a sterile space to reduce introduction of potential contaminants. Use dedicated pipettes, tips, and tubes for sterile work. Pipette up and down gently and throughout protocol to reduce clumps.<br>• Follow closely the Critical steps and tips in **Chapters 5 and 6** (CD8 and CD4 Sample Enrichments)<br>• Load recommended number of cells (30,000 cells per chip) (**Chapter 10**)<br>• Use low protein binding centrifuge tubes |
| Limited frequency of stimulated cells, i.e. those with cytokine signal, on chip *Viability related* | • Leaving thawed cells in DMSO for an extended period<br>• Low viable cells due to low viability input sample and lack of utilization of Ficoll Paque PREMIUM 1.084<br>• Low viable cells due to lack of 2-Mercaptoethanol supplementation to complete RPMI.<br>• Decreased viability due to cell shock | • After thaw, quickly transfer cells from DMSO to complete RPMI to ensure viability of cells<br>• Verify viability of cells is above 80% as stated in **Chapter 4** to ensure protocol is being performed with the highest quality of cells. Use Ficoll Paque PREMIUM 1.084 in **Appendix D2** if viability is less than 80%<br>• Ensure that complete RPMI is supplemented with 2-Mercaptoethanol right before use.<br>• Use reagents at recommended temperatures (i.e. always use warmed media [37°C]) |



Key: ● TIP, ● CRITICAL, ● OPTIONAL

| | | |
|---|---|---|
| Limited frequency of stimulated cells, i.e. those with cytokine signal, on chip *Stimulation step related* | • Recommended anti-CD3e/anti-CD28 stimulation concentration was not used<br>• Recommended anti-CD3e/anti-CD28 stimulation duration was not used | • Use anti-CD3e/anti-CD28 concentrations listed in **Chapter 2** and **Chapter 7**<br>• Use anti-CD3/anti-CD28 timing listed in **Chapter 7**<br>• Use recommended vendor as listed in **Table 3** |